

CERTIFICATE OF ANALYSIS

Lot#: CHM2305-KC-Z

PRODUCT DESCRIPTION

Reference: HuKC

Product: Cryopreserved Human Kupffer Cells

Cellular passage: P0

Size/Quantity: 500.000 cells/vial

Isolation date: 26th April 2023

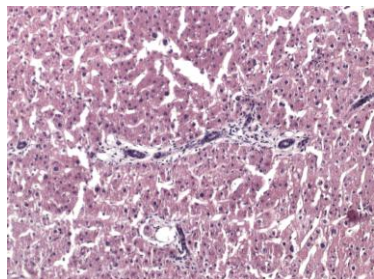
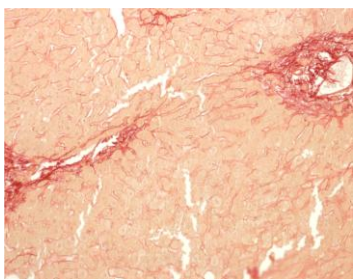
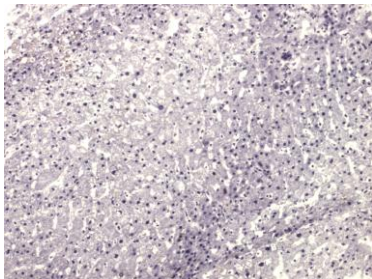
Storage conditions: -196°C using LN₂
Sterility test: Negative for bacteria, yeast, and fungi

DONOR DEMOGRAPHICS

Species	Gender	Race	Age	BMI	Smoker	Alcohol Use	Drug Use
Human	Male	Caucasian	76	24.91	No	No	No
Pathology		Serological Data ¹					
Cholangiocarcinoma		Tested negative less than 3 months before surgery					

Patient informed consent was obtained. ¹The donor was serologically tested negative for following infectious diseases: HIV, Hepatitis B and C, and SARS-CoV-2. Donor medical history was also examined prior to accepting this donor. *For donor's medication information, please contact us.*

DONOR HISTOLOGY

Hematoxylin & Eosin staining	Sirius Red staining	Oil red staining
		

- Hematoxylin & Eosin: Parenchyma with very light microvesicular steatosis present and some detectable necrosis (anuclear hepatocytes) dispersed throughout the parenchyma. Cellular infiltrate in the portal triads close to bile ducts and portal vein

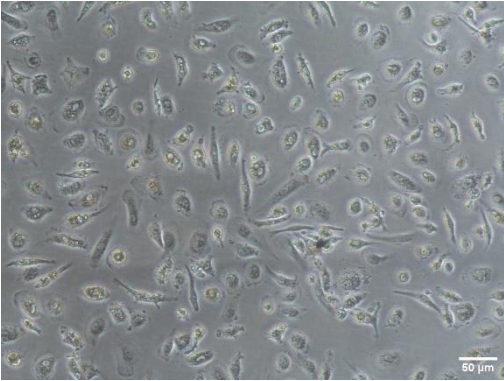
- Sirius red: Liver with light signs of fibrosis, with discrete accumulation of sirius red staining in portal areas and discrete bridging fibrosis. Also, observable matrix deposition in the sinusoidal vessels throughout the parenchyma

- Oil red: Very light vacuolation present in some hepatocytes and negative for oil red staining

Conclusions: Liver with some areas exhibiting light matrix deposition. Most of the tissue devoid of vacuoles and/or fatty accumulation. Cellular infiltrate in the portal areas

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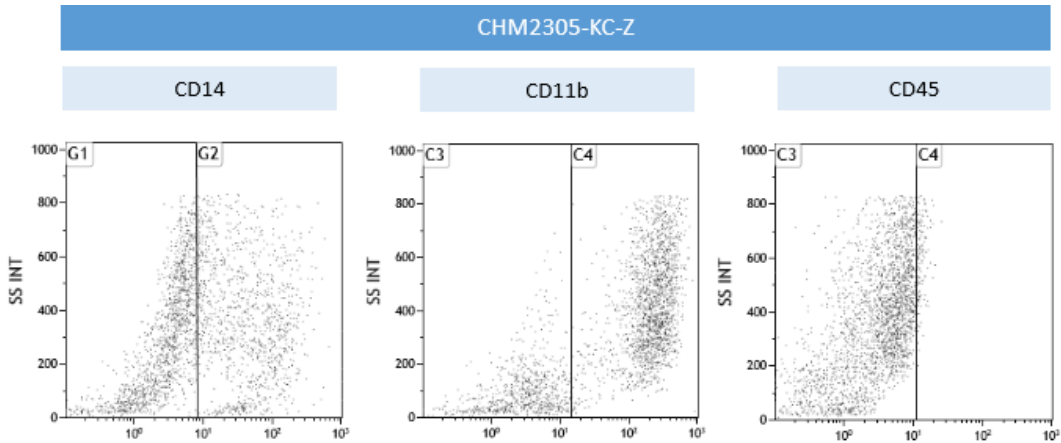
CHARACTERIZATION FOR KUPFFER CELLS

Post Thaw Lot information	
Number of viable cells/vial:	≥ 500.000
Viability (%):	73.29 ± 1.86
Cell seeding density (cells/cm²):	100.000-150.000
Cell morphology 6 days	
	

Kupffer cells were thawed and seeded according to Cytes Biotechnologies protocol. The yield and viability post-thawing was assessed by using the trypan blue exclusion assay. Resuspended human Kupffer cells from post-thaw assessment were plated in 48-well plates in Culture Kupffer Medium. Kupffer cells will begin to attach at approximately day 6, at with point the medium can be replaced.

FLOW CYTOMETRY

Human Kupffer cells cultured for 6 days were analyzed by flow cytometry using specific cell markers.



Flow cytometry of human Kupffer cells. Representative dot-plots of the markers used to characterize human Kupffer cells.

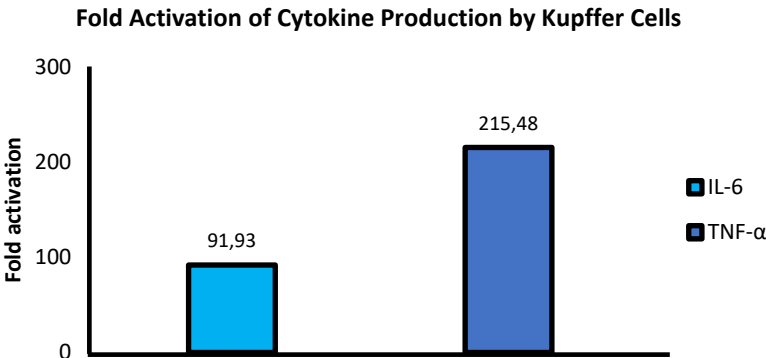
Cell marker	Positive cells
CD14	45.35%
CD11b	71.10%
CD45	N/A

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Flow cytometry analysis from human Kupffer cell population. Cryopreserved human Kupffer cells were thawed and plated in 48-well plates in Culture Kupffer Medium. After 6 days in culture and without any medium replacement, cells were detached and stained with CD14, CD11b, and CD45. Results were expressed in terms of marker expression vs. control (%). Data were obtained by using a Beckman Coulter Gallios Flow Cytometer and were analyzed by using Kaluza Analysis 2.1 software.

LPS STIMULATION

Human Kupffer cells were cultured for 6 days and then, treated with LPS. Fold activation of IL-6 and TNF-α are listed in the graph below.



Fold activation of cytokine production by Kupffer cells. Cryopreserved human Kupffer cells were thawed and plated in 48-well plates in Kupffer Culture Medium. After 6 days in culture and without any medium replacement, cells were treated with 1 ug/ml LPS for 4 hours. At the end of the treatment period, RNA was isolated for mRNA analysis. Results were expressed in terms of fold activation of IL-6 and TNF- α. Data were obtained by using a QuantStudio™ 5 Real-Time PCR System.

If you need help for an experiment, just contact us, our experts will be pleased to assist you

CERTIFICATION:

The viability and performance of the human stellate cells provided depend primarily on the use of appropriate media and reagents, as well as the use of sterile plastics. Likewise, proper handling protocols must be followed. Please note that if these parameters are not carefully considered, the cellular response obtained in the assays may be lower than expected.

Name	Tittle	Signature	Cytes Biotechnologies, S.L.	Date
Pilar Sainz de la Maza	Quality Manager			14/06/23

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