Updated on: 17th June 2024

CERTIFICATE OF ANALYSIS

Lot#: NHM2252-HE-N

PRODUCT DESCRIPTION

Reference: HuHECPMI/4-
Product: Cryopreserved Human Hepatocytes
Category: Plateable, Cytochrome P450 inducible
Spheroid qualified: NO
(see details below: 3D Spheroid formation section)

Isolation date: 6th May 2019
Storage conditions: -196°C using LN_2
Sterility test: negative for mycoplasma, bacteria, yeast, and fungi

DONOR DEMOGRAPHICS

Species	Gender	Race	Age	BMI	HLA	Smoker	Alcohol Use	Drug Use	COD
Human	Male	Caucasian	45	29.1	A11, A68, B35, B53, C04, C04	No	1 drink 3-4x per year	N/A	Anoxia

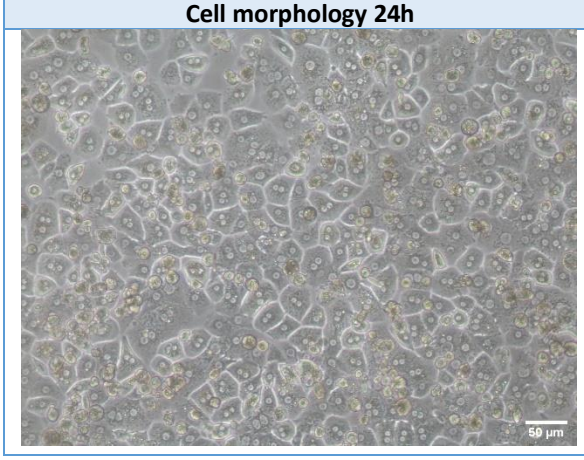
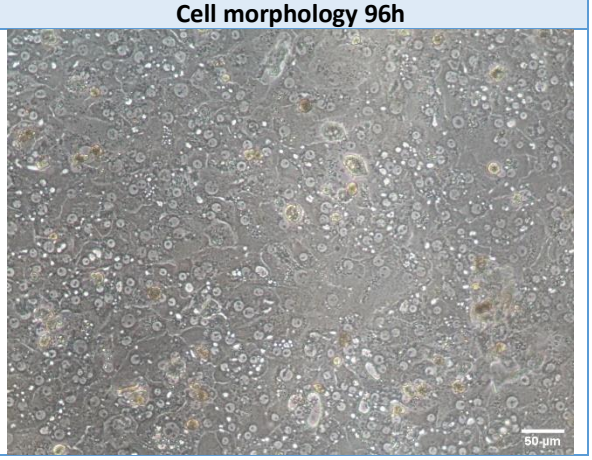
Patient informed consent was obtained. The donor was serologically tested negative for following infectious diseases: HIV, Hepatitis B and C, and syphilis.

CHARACTERIZATION FOR PLATEABLE CELLS

Post Thaw Lot information	Result	SD	n
Number of viable cells (cells/vial):	$5.19x10^6$	± $1.49x10^6$	5
Post-thaw viability (%):	85.478	± 6.28	5
Days in culture after thaw (24w):	9	± 4.24	2
Days in culture after thaw (96w):	4	± 2	2

MONOLAYER ASSESSMENT[1] **Plateable: YES** **Confluence 24h:** 90%

Seeding density in 24 well recommended: $2.12x10^5$ cells/cm^2
Seeding density in 96 well recommended: $2.20x10^5$ cells/cm^2

Cell morphology 24h	Cell morphology 96h
	

Human hepatocytes were thawed and seeded according to BeCytes Biotechnologies culture protocol. The yield and viability were determined by a trypan blue exclusion assay after the thawing process. [1]Resuspended human hepatocytes from post-thaw assessment were plated in collagen-coated 24-well plates in hepatocyte plating medium. Cells were refreshed with hepatocytes maintenance medium during the first change of medium on the day of thawing. Maintenance medium was replaced in the culture every day. If images from the 96-well plates are needed, please contact us.

3D SPHEROID FORMATION

Spheroid morphology
Cytes **does not guarantee** that these primary hepatocytes will be suitable for 3D culture and creation of spheroid structures.

INDUCTION FOR PLATEABLE CELLS

PHASE I: CYP ACTIVITIES EXPRESSED IN pmol/min/mg protein (mean ± SD)

Induction (Specific Activity)				
Enzyme	Basal Activity on day 1	Basal Activity on day 4	Induced Activity on day 4	n-Fold induction
CYP1A2	0.68 ± 0.06	0.78 ± 0.11	6.99 ± 0.40	8.95
CYP2B6	0.78 ± 0.18	0.11 ± 0.01	2.03 ± 0.09	18.60
CYP3A4	0.72 ± 0.12	1.18 ± 0.18	7.98 ± 0.19	6.73

Cryopreserved human hepatocytes were thawed and plated in 24well collagen I coated plates. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. Treatment (n=2 per compound) with vehicle control [0.15% (v/v) DMSO] or inducers (Rifampicin, β-Naphthoflavone and Phenobarbital) began 1-day post-plating and continued for 72 hours. At the end of induction, monolayers were rinsed with PBS and incubated with probe substrate solutions in culture media. See Table 1 for information on each probe substrate. Metabolites were quantified by LC-MS and normalized to protein content. The fold induction was calculated by dividing the induced activity by the vehicle basal activity on the same day in culture.

PHASE I: CYP450 mRNA induction

CYP (mRNA)	n-Fold Induction
CYP1A2	9 ± 1
CYP2B6	24 ± 4
CYP3A4	12 ± 4

Cryopreserved human hepatocytes were thawed, plated in 24well collagen I coated plates in Hepatocyte Plating Medium. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. Maintenance medium was replaced in the cultures daily. Treatment (n=2 per compound) with vehicle control [0.15% (v/v) DMSO] or inducers (Rifampicin, β-Naphthoflavone and Phenobarbital) began 1-day post-plating and continued for 72 hours. At the end of the treatment period, RNA was isolated for mRNA analysis.

Table 1. Substrates Phase I

Enzyme	Probe Substrate	Concentration (µM)	Incubation Time (min)	Metabolite
CYP1A2	Phenacetin	100	30	Acetaminophen
CYP2B6	Bupropion	500	30	Hydroxybupropion
CYP3A4	Midazolam	30	30	1-Hydroxymidazolam

PHASE II: UGTs & SULT ACTIVITIES 24h AFTER SEEDING EXPRESSED IN pmol/min/mg PROTEIN (mean ± SD)

Enzyme	Conjugate	pmol/min/mg
UGT	7-OH coumarin glucuronide	65.70 ± 9.71
SULT	7-OH coumarin sulfate	27.81 ± 2.36

Cryopreserved human hepatocytes were thawed, plated in 24well collagen I coated plates in Hepatocyte Plating Medium. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. On day 1, hepatocytes were incubated with 7-Hydroxycoumarin to assay for UDP-Glucuronosyltransferase (UGT) and Sulfotransferase (SULT) activities. See Table 2 for information on each probe substrate. Metabolites were quantified by LC-MS and normalized to protein content.

Table 2. Substrates Phase II

Enzyme	Probe Substrate	Concentration (µM)	Incubation Time (min)	Metabolite
UGT	7-Hydroxycoumarin	100	30	7-Hydroxycoumarin-glucuronide
SULT	7-Hydroxycoumarin	100	30	7-Hydroxycoumarin-sulfate

If you need help for an experiment, just contact us, our experts will be pleased to assist you

CERTIFICATION:

The viability and performance of the primary human hepatocytes provided depend primarily on the use of appropriate media and reagents, as well as the use of sterile plastics. Likewise, proper handling protocols must be followed. Please note that if these parameters are not carefully considered, the cellular response obtained in the assays may be lower than expected.

Name	Tittle	Signature	Cytes Biotechnologies, S.L.	Date
Pilar Sainz de la Maza	Quality Manager			17/06/24

For basic research use only, not to be used for clinical or diagnostic applications. Products distributed by BeCytes Biotechnologies may contain human material that should be treated as potentially biohazardous.

BeCYTES BIOTECHNOLOGIES

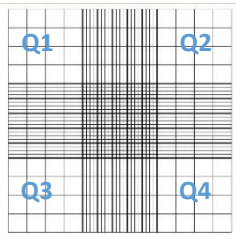
CELL COUNTING

Lot #: _____ **Date:** ____/_____/_____

MORPHOLOGY

❑ Clear cytoplasm	❑ Rounded shape	❑ Cell swelling	❑ Hardly any debris
❑ Clear membranes	❑ Membrane blebbing	❑ Lipid droplets	❑ Prevalent debris

TRYPAN BLUE COUNTING RESULTS



NEUBAUER CHAMBER COUNTING				
Quadrant	Live cells	+	Dead cells	= Total cells
Quadrant 1		+		=
Quadrant 2		+		=
Quadrant 3		+		=
Quadrant 4		+		=
Total		+		=

VIABILITY

$$\frac{(Live\ cells)}{(Total\ cells)} x100 = \qquad Viability\ (\%)$$

YIELD

$$\frac{(Total\ cells) \quad x\ (Dilution\ factor) \quad x\ 10^4\ ^*x\ (Current\ volume) \quad ml}{(Counted\ quadrants)} = \qquad cells\ (Total\ number\ of\ cells)$$

**This factor (10^4) is applicable when it is used a Hemocytometer*

SEEDING DENSITY

$$\frac{(Desired\ number\ of\ cells) \quad cells\ x\ (\ Current\ volume) \quad ml}{(Total\ number\ of\ cells) \quad cells} = \qquad ml\ (Volume\ needed\ for\ your\ cells)$$

Keep in mind the final volume per dish or plate to use (Volume needed) and then calculate the needed volume to add: $(Total\ volume\ well) \quad ml\ -(Cells\ total\ volume) \quad ml = \quad ml\ (Volume\ to\ add)$

Surface of the most common plates for culture:

Brand	24-well plate	96-well plate
ThermoFisher	1.90 cm^2/well	0.32 cm^2/well
Corning®	2.00 cm^2/well	0.36 cm^2/well
Falcon®	1.90 cm^2/well	0.32 cm^2/well
Eppendorf	2.08 cm^2/well	0.37 cm^2/well

COMMENTS

COUNTED BY: