

CERTIFICATE OF ANALYSIS

Lot#: BHum15052

PRODUCT DESCRIPTION

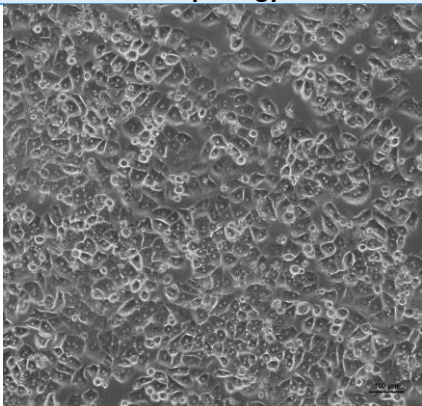
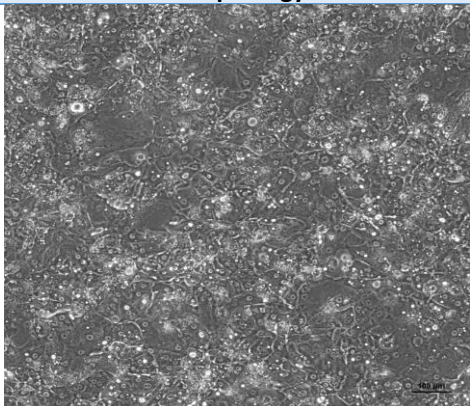
Reference: HuHECPMI/4+**Product:** Cryopreserved Human Hepatocytes**Category:** Plateable, Cytochrome P450 inducible**Spheroid qualified:** NO*(see details below: 3D Spheroid formation section)***Isolation date:** 16th October 2015**Initial Isolation Viability:** 88.00%**Storage conditions:** -196°C using LN2**Sterility test:** negative for bacteria, yeast, and fungi

DONOR DEMOGRAPHICS

Species	Gender	Race	Age	BMI	Smoker	Alcohol Use	Drug Use
Human	Male	Caucasian	54	23	No	No	No
Pathology		Serological Data ¹					
Colorectal cancer hepatic metastases		Tested negative less than 3 months before surgery					

Patient informed consent was obtained. ¹The donor was serologically tested negative for following infectious diseases: HIV, Hepatitis B and C. Donor medical history was also examined prior to accepting this donor. *For donor's medication information, please contact us.*

CHARACTERIZATION FOR PLATEABLE CELLS

Post Thaw Lot information	Result	SD	n
Number of viable cells (cells/vial):	5.03x10 ⁶	±1.69x10 ⁶	5
Post-thaw viability (%):	78.45	± 0.10	5
Days in culture after thaw (24w):	10	± 1.41	4
MONOLAYER ASSESSMENT² Plateable: YES Confluence: 81% Seeding density in 24 well recommended: 2.24x10 ⁵ cells/cm ²			
Cell morphology 24h		Cell morphology 96h	
			

Human hepatocytes were thawed and seeded according to Cytes Biotechnologies culture protocol. The yield and viability were determined by a trypan blue exclusion assay after the thawing process. ²Resuspended human hepatocytes from post-thaw assessment were plated in collagen-coated 24-well plates in hepatocyte plating medium. Cells were refreshed with hepatocytes maintenance medium at first medium during the first change of medium on the day of thawing. Maintenance medium was replaced in the culture every day. If images from the 96-well plates are needed, please contact us.

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3D SPHEROID FORMATION

Spheroid morphology

Cytes **does not guarantee** that these primary hepatocytes will be suitable for 3D culture and creation of spheroid structures while using Cytes protocols.

INDUCTION FOR PLATEABLE CELLS

PHASE I: CYP ACTIVITIES EXPRESSED IN pmol/min/mg protein (mean ± SD)

Enzyme	Induction (Specific Activity)			
	Basal Activity on day 1	Basal Activity on day 4	Induced Activity on day 4	n-Fold induction
CYP1A2	23.81 ± 2.77	0.58 ± 0.13	4.97 ± 5.09	8.53
CYP2B6	7.37 ± 1.53	0.92 ± 0.14	1.97 ± 0.25	2.14
CYP3A4	16.06 ± 3.27	0.82 ± 0.23	2.92 ± 0.71	3.57

Cryopreserved human hepatocytes were thawed and plated in 24well collagen I coated plates. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. Treatment (n=2 per compound) with vehicle control [0.15% (v/v) DMSO] or inducers (Rifampicin, β-Naphthoflavone and Phenobarbital) began 1-day post-plating and continued for 72 hours. At the end of induction, monolayers were rinsed with PBS and incubated with probe substrate solutions in culture media. See Table 1 for information on each probe substrate. Metabolites were quantified by LC-MS and normalized to protein content. The fold induction was calculated by dividing the induced activity by the vehicle basal activity on the same day in culture.

PHASE I: CYP450 mRNA induction

CYP (mRNA)	n-Fold Induction
CYP1A2	19
CYP2B6	19
CYP3A4	23

Cryopreserved human hepatocytes were thawed, plated in 24well collagen I coated plates in Hepatocyte Plating Medium. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. Maintenance medium was replaced in the cultures daily. Treatment (n=2 per compound) with vehicle control [0.15% (v/v) DMSO] or inducers (Rifampicin, β-Naphthoflavone and Phenobarbital) began 1-day post-plating and continued for 72 hours. At the end of the treatment period, RNA was isolated for mRNA analysis.

Table 1. Substrates Phase I

Enzyme	Probe Substrate	Concentration (μM)	Incubation Time (min)	Metabolite
CYP1A2	Phenacetin	100	30	Acetaminophen
CYP2B6	Bupropion	500	30	Hydroxybupropion
CYP3A4	Midazolam	30	30	1-Hydroxymidazolam

PHASE II: UGTs & SULT ACTIVITIES 24h AFTER SEEDING EXPRESSED IN pmol/min/mg PROTEIN (mean ± SD)

Enzyme	Conjugate	pmol/min/mg
UGT	7-OH coumarin glucuronide	375.66 ± 46.26
SULT	7-OH coumarin sulfate	20.70 ± 1.21

Cryopreserved human hepatocytes were thawed, plated in 24well collagen I coated plates in Hepatocyte Plating Medium. Cells were overlaid with Matrigel® (Corning) in Human Hepatocyte Maintenance Medium at first medium change at day of thawing. On day 1, hepatocytes were incubated with 7-Hydroxycoumarin to assay for UDP-Glucuronosyltransferase (UGT) and Sulfotransferase (SULT) activities. See Table 2 for information on each probe substrate. Metabolites were quantified by LC-MS and normalized to protein content.

Table 2. Substrates Phase II



Enzyme	Probe Substrate	Concentration (μM)	Incubation Time (min)	Metabolite
UGT	7-Hydroxycoumarin	100	30	7-Hydroxycoumarin-glucuronide
SULT	7-Hydroxycoumarin	100	30	7-Hydroxycoumarin-sulfate

If you need help for an experiment, just contact us, our experts will be pleased to assist you

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CERTIFICATION:

The viability and performance of the primary human hepatocytes provided depend primarily on the use of appropriate media and reagents, as well as the use of sterile plastics. Likewise, proper handling protocols must be followed. Please note that if these parameters are not carefully considered, the cellular response obtained in the assays may be lower than expected.

Name	Tittle	Signature	Cytes Biotechnologies, S.L.	Date
Pilar Sainz de la Maza	Quality Manager			24/03/23

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CELL COUNTING

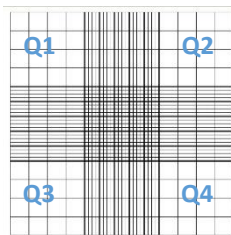
Lot #: _____

Date: ____/____/____

MORPHOLOGY

- ☐ Clear cytoplasm ☐ Rounded shape ☐ Cell swelling ☐ Hardly any debris
☐ Clear membranes ☐ Membrane blebbing ☐ Lipid droplets ☐ Prevalent debris

TRYPAN BLUE COUNTING RESULTS



NEUBAUER CHAMBER COUNTING					
Quadrant	Live cells	+	Dead cells	=	Total cells
Quadrant 1		+		=	
Quadrant 2		+		=	
Quadrant 3		+		=	
Quadrant 4		+		=	
Total		+		=	

VIABILITY

$$\frac{(\text{Live cells})}{(\text{Total cells})} \times 100 = \text{Viability (\%)}$$

YIELD

$$\frac{(\text{Total cells}) \times (\text{Dilution factor}) \times 10^4 \times (\text{Current volume})}{(\text{Counted quadrants})} \times \frac{\text{ml}}{\text{cells}} = \text{cells (Total number of cells)}$$

*This factor (10⁴) is applicable when it is used a Hemocytometer

SEEDING DENSITY

$$\frac{(\text{Desired number of cells})}{(\text{Total number of cells})} \times \frac{\text{cells} \times (\text{Current volume})}{\text{cells}} \times \frac{\text{ml}}{\text{cells}} = \text{ml (Volume needed for your cells)}$$

Keep in mind the final volume per dish or plate to use (Volume needed) and then calculate the needed volume to add: $(\text{Total volume well}) \text{ ml} - (\text{Cells total volume}) \text{ ml} = \text{ml (Volume to add)}$

Surface of the most common plates for culture:

Brand	24-well plate	96-well plate
ThermoFisher	1.90 cm ² /well	0.32 cm ² /well
Corning®	2.00 cm ² /well	0.36 cm ² /well
Falcon®	1.90 cm ² /well	0.32 cm ² /well
Eppendorf	2.08 cm ² /well	0.37 cm ² /well

COMMENTS

COUNTED BY:

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